Separation of Bile Acid Isomers Based on Differential Mobility Spectrometry

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ABSTRACT

Bile acids are involved in a wide range of biological functions including lipid resorption, immunological functions and metabolic regulation. Through metabolic transformations, isomeric and isobaric variants are generated, which makes the unequivocal identification and quantification of individual chemical species difficult.

Here, differential ion mobility spectrometry (DMS, SelexION[®] Technology) was used as a methodology for the separation of bile-acid isomers. SelexION[®] device is an ion mobility technology which separates molecules based on their dipole moment instead of m/z. This separation was used in conjunction with liquid chromatographic separation (LC-DMS-MS) as well as with direct infusion (DMS-MS). While the combination with chromatographic separation may improve selectivity, the separation power of SelexION[®] technology is sufficient for a clear separation of isomers, enhancing the selectivity of the measurement, and also allowing for infusion-based fast quantification without the need for LC separation



Figure 1. High Selectivity Quantification using SelexION[®] Technology on the QTRAP[®] 6500 LC-**MS/MS System.** The SelexION[®] Technology is an easy to install differential mobility separation (DMS) device that can be installed on a QTRAP® 5500 or 6500/6500+ LC-MS/MS system, attaching in front of the curtain plate (Left). Gas draws the ions towards the orifice (Middle) while a Separation Voltage (SV) in the form of an asymmetric waveform is applied to the flat plates, which alternates between high field, K(E) and low field, K(0). This moves the charged ion back and forth between plates, an ion will have net drift base on its high and low field mobility. A separation voltage (SV) is applied as the filtering voltage and the compensation voltage (CoV, a small DC offset between the plates) is applied as the restoring voltage, which can be tuned for the compound of interest. Other co-eluting nearly isobaric species that tune with different compensation voltages will be filtered away (right).

Methods

Sample Preparation: Commercially available bile acid standards were prepared in 1 mM in 10% DMSO as stock solution. Two groups of bile acid standards were used: taurodeoxycholic acid (TDCA), taurochenodeoxycholic acid (TCDCA), tauroursodeoxycholic acid (TUDCA) with monoisotopic mass of 499.3 Da and formula C₂₆H₄₅O₆NS and glycodeoxycholic acid (GDCA), glycochenodeoxycholic acid (GCDCA) and glycoursodeoxycholic acid (GUDCA) with monoisotopic mass of 449.3 Da and formula $C_{26}H_{43}O_5N$. For direct infusion, compounds were prepared in 50% MeOH at 0.1 µM concentration.

Liquid Chromatography: For LC separation compounds were prepared in 50% MEOH at 10 nM concentration was done on a SCIEX ExionLC[™] AD HPLC system. . The samples were analyzed on a QTRAP[®] 6500+ LC-MS/MS system equipped with a SelexION[®] device.

Results

Using the SelexION[®] device, the bile acid standards were infused individually and as mixtures to determine the CoV values of the different isomers. As shown in Figure 3, the species of the two groups were infused and the compensation voltage ramped over a range of -40V to 30V. The total ionogram shows the presence of a number of separate peaks.

The LC-DMS-MS analysis were also performed. The addition of DMS to LC separation has several advantages:

Figure 2. Bile acids are derivatives of cholesterol, which carry a short carboxylic acid chain on the **C17** atom of the cholesterol back bone. A range of modifications can occur on the carboxylic function, which define the class of the bile acid. Two groups of isobaric bile acids were analyzed by SelexION[®] Technology: 1) Taurocholic acid (left), a conjugate of bile acid and taurine and 2) Glycocholic acid, which is a conjugate of bile acid with glycine. The isomers within each of the two classes differ only with respect to the location or configuration of a hydroxyl group. In the DCA-Variant, the hydroxyl group is located on the C-12 of the cholesterol skeleton, while in the CDCA and the UDCA variants, the hydroxyl group is located on the C-7 atom in alpha- or in beta-configuration, respectively.

Next, LC-DMS-MS analysis was performed. The addition of DMS to the LC separation has several advantages: including the removal of chemical background for better quantification, less requirement for development of chromatographic separation, no requirement for retention time assignment and less reliance on compound specific fragments which may be low abundant.

As show in Figure 4, the same transition was used, but with the addition of DMS, specific isomers can be determined by use of the respective specific compensation voltages.

- removal of chemical background for better quantification
- less requirement effort for development of chromatographic separation
- there is no requirement for retention time assignment
- less reliance on compound specific fragments which may be low abundant.







corresponding CoV to enter the MS for detection.

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