

High-Throughput and Automated Software Workflow Strategies for Small Molecule Identification and Characterization Using a High Resolution Q-TOF Mass Spectrometer



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ABSTRACT

Transformation of accurate mass product spectra into putative structures of metabolites poses a bottleneck in early discovery studies to identify metabolic soft spots, as well as in the subsequent characterization of active metabolites. Here we present an automated software workflow for the proposal of structure analogues and its application to a variety of small molecules. Attributes affecting performance of this qualitative workflow are discussed.

INTRODUCTION

While software-predicted results for metabolite structure proposals cannot beat human expertise, software strategies can greatly aid in streamlining the process of metabolite soft-spot identification.

In this work, a high-throughput and completely automated workflow strategy is investigated for small molecule metabolite identification and characterization using a high-resolution Q-TOF mass spectrometer.

Both parent-drug tailored IDA and generic SWATH® acquisition methods have been used for data collection. The resulting files were interrogated to find drug-related material. Since the MS/MS data were collected in parallel with accurate TOF MS data, metabolite fragments in conjunction with the known parent drug structure provided grounds for ranking of putative structures and proposal of biotransformation reactions.

MATERIALS AND METHODS

10 µM microsomal incubations of various compounds (nefazodone, propranolol, bromocriptine, diclofenac, verapamil, imipramine) were analyzed using a TripleTOF® 5600 system coupled with a Shimadzu Prominence UFLC System. A 300 µL/min gradient was utilized with a Kinetex C18 (2.6 µ) 2 *50 mm column and run for 10 minutes.

Table 1. Core MS data collection parameters for variety of acquisition strategies

	IDA positive	SWATH positive	IDA negative	SWATH negative
TOF MS mass range	100 - 1000	100 - 1000	100 - 1000	100 - 1000
TOF MS/MS mass range	100 - 1000	100 - 1000	100 - 1000	100 - 1000
TOF MS/MS CE	35 ± 15	25 to 55	35 ± 15	-4 to -40
SWATH Q1 coverage	N/A	300 - 800 (25*20)	N/A	100 - 915 (variable)
Number of MS/MS experiments	3	20	3	20
Cycle time (ms)	390	900	390	600

Resulting LC/MS data was mined with MetabolitePilot™ 2.0 software using both hypothesis-driven and generic strategies to find drug-related material. Sites of modifications for major metabolites were proposed, ranked and scored within the batch processing. Automatically generated proposed metabolite structures were then validated either manually or against published data.

DISCUSSION

Within MetabolitePilot 2.0 software structures are proposed for the following metabolite types:

- One or more dealkylation cleavage,
- One biotransformation,
- Combination of cleavage and biotransformation.

Scoring and ranking of potential modification sites is based on projection of information from the annotated MS/MS spectrum of parent drug onto the metabolite MS/MS spectrum. Subset of SOM candidate atoms is retrieved from the biotransformation database. Score for each SOM candidate atom is derived from the evidence supporting its both unchanged and modified states. Proposed structures are generated by attaching/removing biotransformation structure motif onto parent drug.

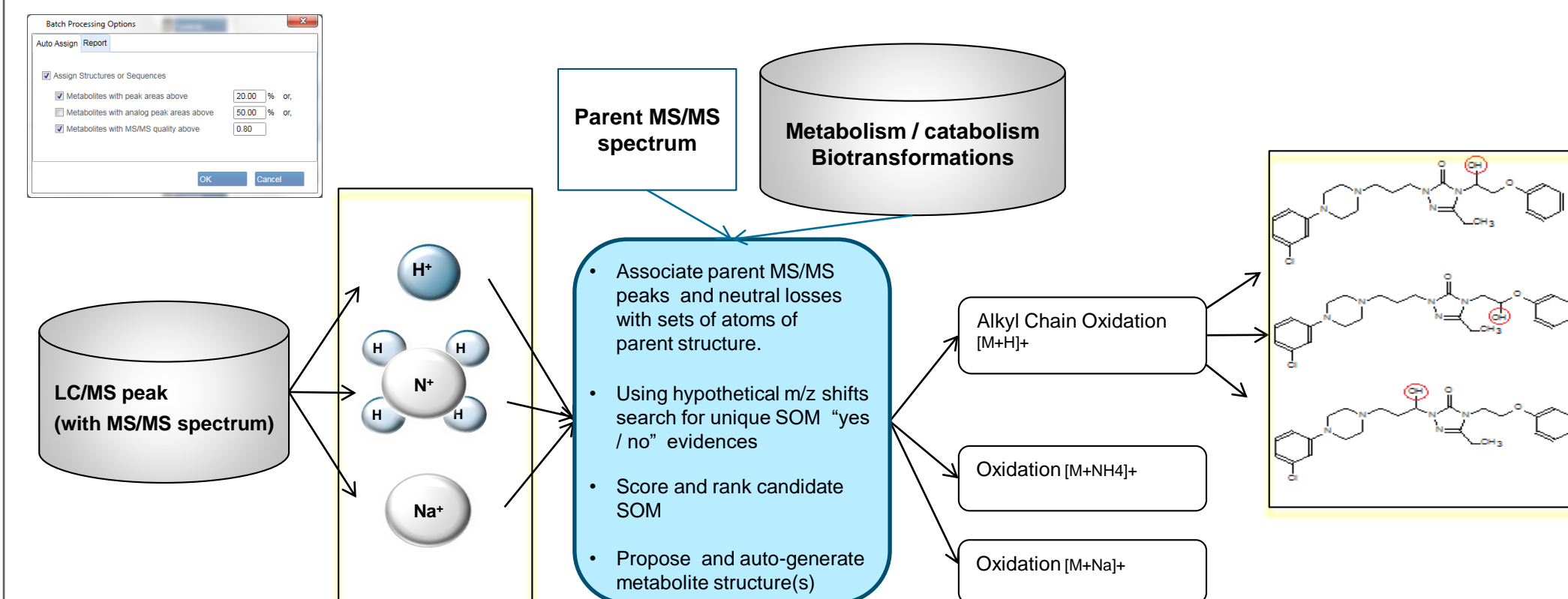
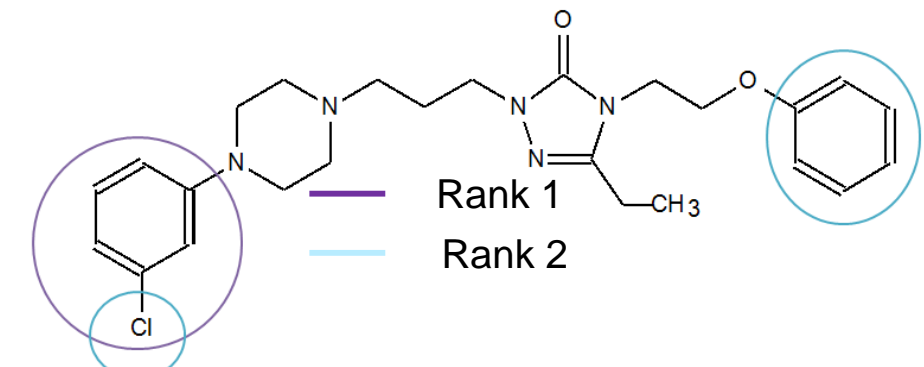


Figure 1. Automated structure proposal within an LC / HRMS qualitative data processing pipeline.

Accurate mass MS and MS/MS data were collected in parallel; complementary ions were grouped in TOF MS and MS/MS spectra of protonated / deprotonated precursor were used for structure proposal.

RESULTS

Example 1: Nefazodone dealkylation metabolite, Loss of C₆H₃Cl, m/z 360.2392, RT 2.09 minutes

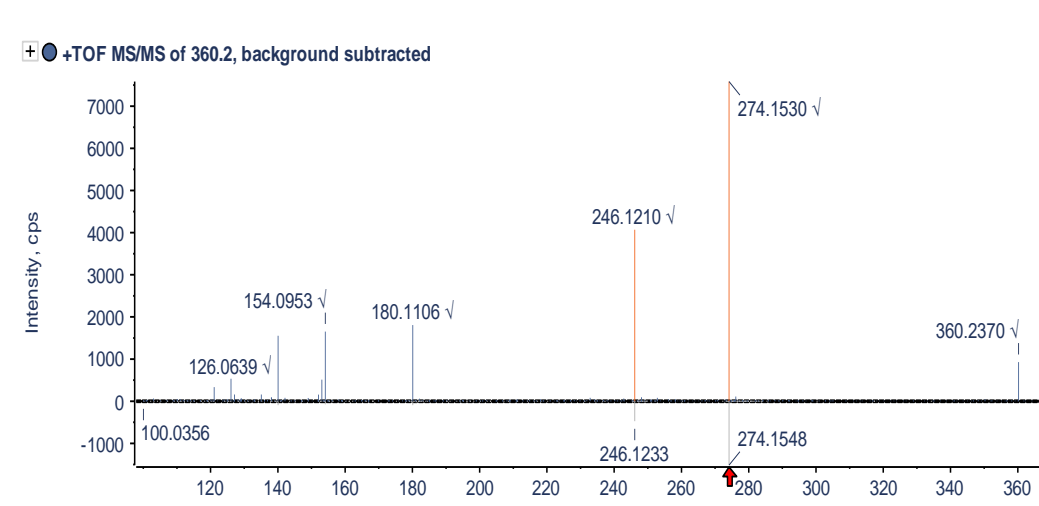


Assigned: 23 of 30 peaks, score for 23 proposed assignments: 534.0

Fragments: 17 of 43 Proposed Formulae

Use	Mass (m/z)	Ion Formula	Error (mDa)	Intensity (cps)	RDB	Proposed Structures	Score
1	110.0843	C7H12N	-2.2	48.3	3.0	1	32.0
2	121.0622	C8H9O	-2.6	335.7	5.0	1	40.0
6	138.1005	C7H12N3	-2.1	80.6	4.0	1	33.0
12	180.1106	C9H14N3O	-2.5	1808.3	5.0	1	36.0
15	274.1530	C15H20N3O2	-2.0	7583.3	8.0	1	40.0
17	360.2370	C19H25N3O2	-2.4	929.1	8.0	1	44.0
18	149.0201	C8H9NO2	-1.9	79.1	7.5	2	5.0
4	246.1210	C13H18N3O2	-2.7	4066.6	8.0	3	22.0
16	276.1601	C14H20N3O2	2.0	112.2	7.5	3	8.5
9	152.0800	C7H10N3O	-1.9	156.9	5.0	6	36.0
10	153.0881	C7H11N3O	-1.5	511.9	4.5	6	33.5
11	154.0853	C7H12N3O	-2.2	1652.5	4.0	6	36.5
3	124.0496	C5H8N3O	-1.0	48.3	5.0	8	21.0
4	126.0639	C5H9N3O	-2.3	535.3	4.0	8	21.5
5	138.0633	C6H9N3O	-2.9	96.6	5.0	8	36.0
7	140.0795	C6H10N3O	-1.9	1552.7	4.0	8	36.5
13	211.1535	C10H18N4O	-1.8	48.2	4.0	9	32.5

Two "Loss of C₆H₃Cl" structures were considered.



Assigned: 23 of 30 peaks, score for 23 proposed assignments: 447.0

Fragments: 13 of 43 Proposed Formulae

Use	Mass (m/z)	Ion Formula	Error (mDa)	Intensity (cps)	RDB	Proposed Structures	Score
1	136.0790	C7H9N3	-0.1	160.4	5.5	1	31.0
2	138.1005	C7H12N3	-2.1	80.6	4.0	1	32.0
3	180.1106	C9H14N3O	-2.5	1808.3	5.0	1	36.0
4	360.2370	C19H25N3O2	-2.4	929.1	8.0	1	44.0
5	110.0843	C7H12N	-2.2	48.3	3.0	3	32.0
6	152.0800	C7H10N3O	-1.9	156.9	5.0	7	36.0
7	153.0881	C7H11N3O	-1.5	511.9	4.5	7	33.5
8	154.0853	C7H12N3O	-2.2	1652.5	4.0	7	36.5
9	211.1535	C10H18N4O	-1.8	48.2	4.0	7	8.5
10	124.0496	C5H8N3O	-1.0	48.3	5.0	9	29.0
11	126.0639	C5H9N3O	-2.3	535.3	4.0	9	32.0
12	138.0633	C6H9N3O	-2.9	96.6	5.0	9	36.0
13	140.0795	C6H10N3O	-1.9	1552.7	4.0	9	36.5

Proposed structure 1 is supported by 6 unique fragment structures including one for fragment 274.1530. No unique fragments support attachment of phenyl group to piperazine ring.

MS/MS Collection	Compound Name	Metabolite Name	RT (min)	m/z	Polarity	Candidate Structures	Structure Count* (Generated/Expected)	Rank	Normalized Score (%)	NOTES
IDA	Simvastatin	Oxidation	4.48	435.2747	+	17	2 / 2 ^[1]	1	100	
IDA	Simvastatin	Demethylation	4.79	405.2639	+	0	0 / 0	1	100	
IDA	Simvastatin	Oxidation	6.08	258.0967	+	11	1 / 1 ^[2]	1		
IDA	Methocarbamol	Oxidation	7.27	258.0972	+	11	1 / 1 ^[2]	1	100	
IDA	Verapamil	Loss of CHNO + Demethylation	8.03	185.0802	+	1	manual interpretation	n/a		
IDA	Verapamil	Loss of C10H12O2 and CH2 and Oxidation	2.49	293.1858	+	36	manual interpretation	n/a		
IDA	Verapamil	Loss of CH2 and CH2	3.03	427.2592	+	10	1 / 1 ^[3]	1	100	
IDA	Diclofenac	Oxidation	1.4	310.0039	-	13	4 / 4 ^[4]	1	100	
IDA	Benzbromarone	Oxidation	9.21	436.9021	-	10	2 / 2 ^[5]	1	100	
IDA	Bromocriptine	Oxidation	5.7	668.2154	-	22	1 / 1 ^[6]	1	100	
SWATH	Propranolol	Oxidation	1.87	276.1595	+	6	3 / 3 ^[7]	1 to 2	100/94.3	
SWATH	Propranolol	Oxidation	2.03	276.16	+	6	3 / 3 ^[7]	1 to 2	100/97.3	
SWATH	Propranolol	Glucuronidation	2.52	436.1979	+	2	1 / 1 ^[7]	1	100	
SWATH	Propranolol	Glucuronidation	2.69	436.1991	+	2	1 / 1 ^[7]	1	100	
SWATH	Propranolol	Loss of C3H7N+Demethylation to Carboxylic Acid	3.24	233.0822	+	1	1 / 1 ^[7]	1	100	
SWATH	Nefazodone	Loss of C10H11CIN2+Ketone Formation	2.66	290.1503	+	7	0 / 1 ^[8]			*
SWATH	Nefazodone	Ethyl to Alcohol	3.43	458.1951	+	1	0 / 1 ^[8]			**
SWATH	Nefazodone	Loss of C6H3Cl	2.09	360.2392	+	2	1 / 1 ^[8]	1	100	
SWATH	Nefazodone	Loss of C6H4	2.44	394.2006	+	1	1 / 1 ^[8]	1	100	
SWATH	Nefazodone	Loss of C15H19N3O2+Oxidation	2.73	213.0784	+	11	1 / 1 ^[8]	1	100	
SWATH	Nefazodone	Loss of C15H19N3O2+Oxidation	0.57	213.0791	+	11	1 / 1 ^[8]	1	100	
SWATH	Nefazodone	Loss of C6H3Cl+Oxidation	1.69	376.2343	+	38	12 / 2 ^[8]	6	99.3/98.3	
SWATH	Nefazodone	Loss of C10H11CIN2+Oxidation	3.24	292.1657	+	15	1 / 1 ^[8]	1	100	
SWATH	Nefazodone	Loss of C6H3Cl+Ketone Formation	2.19	374.2179	+	18	2 / 1 ^[8]	2	99.7	
SWATH	Nefazodone	Loss of C6H4+Oxidation	2.17	410.1948	+	20	No reference found	n/a		
SWATH	Nefazodone	Loss of C10H11CIN2+Demethylation to Carboxylic Acid	3.35	306.1445	+	2	1 / 1 ^[8]	1	100	
SWATH	Diclofenac	Oxidation	1.32	310.0044	-	13	4 / 4 ^[4]	1	100	
SWATH	Midazolam	Demethylation to Carboxylic Acid	3.67	356.0591	+	1	No reference found	n/a		
SWATH	Midazolam	Oxidation	2.55	342.081	+	14	2 / 2 ^[9]	1	100	
SWATH	Bromocriptine	Oxidation	3.9	670.2254	+	22	2 / 2 ^[9]	1	100	

* Loss of C10H11CIN2, +O, -2H (complex 3-step biotransformation, not in automated workflow scope)
** oxidation and re-arrangement (currently not represented in biotransformation database)

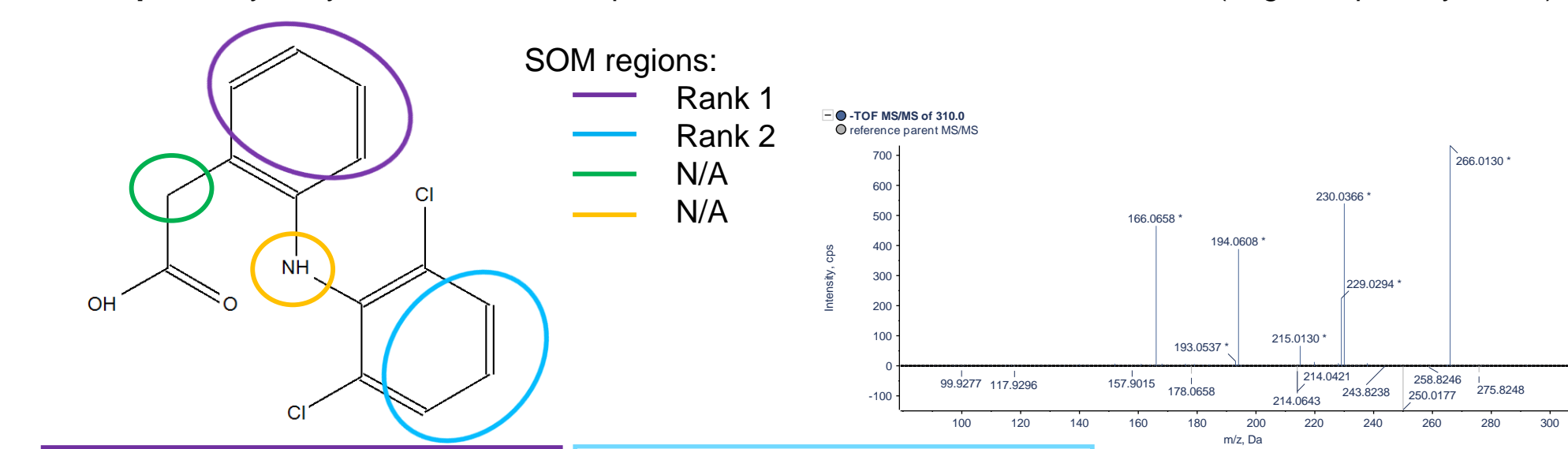
Table 2. Validation of automated structure proposal workflow.

Within the workflow validation, we found that low m/z fragments that originated from a limited specific region of the molecule provided confident insight into structure features. Also, informative MS/MS spectrum of parent drug that covered all portions of parent molecule typically enhanced contrast in candidate structure scores and confidence in putative structure assignment.

The new structure proposal and ranking workflow is complementary to existing "Interpret" functionality in MetabolitePilot™ software. The processing workflow as indicated in Figure 1 is expandable to additional cleavage and biotransformation combinations.

Compatible parameters are critical for MS/MS fragmentation annotation, in examples 1 and 2 up to 30 fragments with S/N above 3 were used, mass tolerance was 5mDa, and in-silico fragments were generated by breaking up to 4 bonds (including aromatic ring) of the parent molecule.

Example 2: Hydroxy metabolite of Diclofenac, m/z = 310.0039, RT = 1.4 minutes (negative polarity mode)



Assigned: 16 of 30 peaks, score for 16 proposed assignments: 521.0

Fragments: 14 of 23 Proposed Formulae

Use	Mass (m/z)	Ion Formula	Error (mDa)	Intensity (cps)	RDB	Proposed Structures	Score
1	152.0512	C8H9O3	-3.3	7.0	4.5	1	48.5
2	164.0518	C12H9N	1.2	4.0	10.0	1	30.5
3	166.0659	C13H9N	-0.5	465.5	9.0	1	38.5
4	176.0505	C13H9N	-0.1	7.0	11.0	1	38.5
5	179.0583	C13H9NO	-1.4	8.0	10.0	1	31.0
6	193.0537	C13H9NO	0.4	15.4	10.0	1	38.0
7	194.0608	C13H9NO	-0.4	388.2	10.0	1	39.5
8	194.0798	C13H9NO	2.0	3.9	8.0	3	34.5
9	143.0464	C7H9O3	1.5	5.5	3.5	4	33.0
10	228.0209	C13H9NO2	-1.2	8.8	10.0	6	38.0
11	229.0284	C13H9NO2	-0.6	224.5	9.5	6	40.0
12	230.0366	C13H9NO2	-1.3	540.0	9.0	6	38.5
13	266.0130	C13H9NO2NO	-1.5	732.3	8.0	8	42.5
14	215.0130	C13H9NO2	-1.3	66.3	9.5	22	38.0

CONCLUSIONS

A fully automated LC HRMS workflow involving either IDA or SWATH® collection and a comprehensive LC/MS data processing including metabolite structure proposal, was demonstrated and validated using variety of model compounds.

This workflow integrated within MetabolitePilot™ 2.0 software, streamlines data processing and enables routine soft-spot analysis capturing major metabolites.

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Document number: RUO-MKT-10-5828-A